

Purpose:

To demonstrate voltage clamp experiment on a CNS neuron containing Na and Ca channels.

General Information:

Program name = "CNS vclamp.rt"

Location = Any of the Power Macintosh computers in the MSL

Directions:

- 1) Double click on the "CNS vclamp.rt" icon to load the program.
- 2) Click on the ARROW button or select "run" from the "operate" menu to begin execution. You may **let the program continue to run** throughout the exercise.
- 3) Any of the parameters (e.g. the stimulator or ionic concentrations) can be altered "on the fly" to test their effects on the currents.
- 4) To reset all parameters to their default values, go to the "operate" menu and select "reinitialize all to default".

Problems:

1) Characteristics of voltage-gated sodium, calcium and potassium currents.

a) Start the program. The default values show the currents elicited following a rapid voltage step from -70 to -20 mV. Compare and contrast the direction and **time course** of the Na⁺, K⁺, Ca²⁺ currents:

Explain why iNa returns to zero within 10 ms while iK remains elevated: _____

Why is the time course faster than for iCa? _____

Now turn "Trace Memory" ON, then set V_{step} to +10 mV. This allows a direct comparison of the traces obtained at -20 mV (white lines) with those at +10 mV (colored lines). How are the iNa, iCa and iK traces at these two voltages similar? _____ How are they different? _____

b) Reset all variables so that V_{step} = -20 mV. Turn "Trace Memory" ON and set V_{step} = +50 mV. Explain why:

gNa is now larger but iNa is smaller: _____

iCa is now larger: _____

gK is now larger: _____

iK is now larger: _____

c) Now increase V_{step} in 1 mV increments from +50 to +80 mV while observing iNa. Describe what happens to Na⁺ current: _____

At what value of V_{step} does iNa change direction? _____ (= reversal potential). You should have been able to calculate this number in advance; how? _____. Set V_{step} = +20 mV and alter the reversal potential for Na⁺ using the Na_{out} control. What is the [Na]_{out} which makes E_{rev} for Na⁺ = +20 mV? _____.

Repeat the above procedure for iCa (if you can). E_{rev} for iCa = _____. What is the [Ca]_{out} which makes E_{rev} for Ca⁺⁺ = +20 mV? _____.

d) Reset all variables. To determine the **threshold** for activation of iNa, set V_{step} to -70 mV and increase it in 1 mV increments until Na⁺ current just begins to activate. What is the potential at which this happens? _____. Repeat this procedure while observing iK (or gK) and determine the activation threshold for K⁺ current: _____. Repeat this procedure while observing iCa (or gCa) and determine the activation threshold for Ca⁺⁺ current: _____. Why would this be different from Na⁺? _____

_____.

2) Effects of changing ion concentrations

a) Reset all variables. Set Na_{in} to 140 mM. What will be the new value of the Na^+ equilibrium potential? _____. How could this be verified experimentally? (try it) _____.
Do g_{Na} , g_K or i_K change with changes in Na_i ? ____ Why or why not? _____

Do i_{Na} or i_{Tot} change with changes in Na_i ? ____ Why or why not? _____

How do changes in internal or external Ca^{++} affect i_{Na} ? _____.

(In a real axon preparation, increases in external Ca^{++} would shift the activation threshold for Na^+ channels).

How do changes in internal or Ca^{++} affect i_{Ca} ? _____ (the changes occur in the same direction in a real cell but result from a different process).

3) Effects of selective blockade.

a) Reset all variables. Now look only at the i_{Tot} curve. Note the complex shape of i_{Tot} as the membrane is progressively depolarized. Sketch below the family of i_{Tot} curves one would obtain if V_{step} were progressively increased from -30 mV to +90 mV in 30 mV steps.

This should look similar to diagrams shown in most textbooks. Why does i_{Tot} have a slight "hump" at 1 ms when $V_{step} = +100mV$? _____

b) Set $V_{step} = 0$ mV and turn Trace Memory ON. Lower Na_{out} from 140 to 14 mM and note the effect on i_{Tot} : _____.

This will simulate an experiment in which external Na^+ is substituted with an impermeable ion .

A second way to eliminate inward current is to block Na^+ channels with tetrodotoxin (TTX). Set Na_{out} back to 140 mM and increase [TTX] to "hi". Record the effect : _____.

A third way to eliminate Na^+ current is to clamp the membrane at E_{Na} so that no Na^+ current flows (you already did this in #1c).

c) Reset all variables. Set V_{step} to 0 mV and turn Trace Memory ON. Now lower K_{in} from 140 to 5 mM; this simulates an experiment in which K^+ inside the axon is substituted with a less permeable ion such as Cs^+ . Note the effect on i_{Tot} : _____.

A second way to eliminate outward current is to block K^+ channels with tetraethylammonium (TEA). Set K_{in} back to 140 mM and increase [TEA] to "hi". Record the effect : _____.

A third way to eliminate K^+ current is to clamp the membrane at E_K so that no K^+ current flows . Why is this a useless experiment ? _____.

d) Reset all variables. Set V_{step} to 0 mV and turn Trace Memory ON. Eliminate inward Ca current using the dihydropyridine channel blocker nifedipine. Record the effect on i_{Tot} : _____.

Keep [nifedipine] hi and increase [TTX]; explain the change in i_{Tot} : _____.